



The Complementary Effects of Salicylic Acid, Alginic Acid, Chitosan and Humic Acid to Improve the Yield and Bioactive Component of Echinacea Plants

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Abstract

The present research sought to evaluate the impacts of various treatments via foliar application with some bio-stimulants (i.e. salicylic acid (200 & 400 mg/L.), alginic acid (250 and 500 mgL⁻¹) as well as chitosan (250 & 500 mgL⁻¹) and humic acid at 3kg/Fadden soil addition compared with control (Distilled water) on flowering parameters (start of inflorescence anthesis (days), inflorescences number/ plant, dry weight of inflorescences, main inflorescence height, main inflorescence diameter and dry weight of main inflorescence /plant) also, some bio-active compounds content (caffeic acid derivatives plus total alkamides) of (*Echinacea purpurea*) plants grown in normal conditions. The results clearly show that all treatments significantly enhanced all studied flowering parameters and some bio-active compounds in echinacea plants compared with control in both seasons. In addition, foliar application treatments of SA at 200 mg L⁻¹ also, chitosan at 500 mgL⁻¹ either alone or in conjunction with humic acid soil addition considerably boosted all the above mentioned flowering parameters. As for some bio-active compound content (caffeic acid derivatives and total alkamides) compared with the other treatments in both seasons, salicylic acid at 200 mg L⁻¹ with humic acid as soil addition was the best successful treatment in this respect. Generally, the foliar spraying treatments by 200mgL⁻¹ of salicylic acid and chitosan (500 mgL⁻¹) in combination with humic acid soil addition were the best influential treatments for enhancing flowering traits and some bio-active substances contant of echinacea plants in normal environment, as well as we recommended used of these treatments.at the same field condition.

Keywords: *Echinacea purpurea* L., flowering parameters, caffeic acid derivatives, total alkamides, growth bio-stimulants, salicylic acid, alginic acid, chitosan, humic acid.

1. Introduction

Purple coneflower (*Echinacea purpurea* L.) (EP) is a herbaceous perennial plant belong to the Asteraceae family. EP is highly appreciated for its therapeutic, economic as well as ecological importance. It has drawn notice because its immune-boosting so anti-inflammatory Anti-inflammatory features, making it a crucial resource in both traditional and modern medicine. (Gupta *et al.*, 2012 and Salgado-Salazar *et al.*, 2023).

Echinacea purpurea is renowned for its immunomodulatory effects, containing active materials as alkylamides, glycoproteins, polysaccharides as well as derivatives of caffeic acid, which enhance immune system activity and aid in the prevention and therapy of illnesses like viral rhinitis so influeinza (Coelho *et al.*, 2020 and Burlou-Nagy *et al.*, 2022). Recent studies confirm the plant's safety and effi-cacy, with research showing a favorable safety profile and effective

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pharmacokinetics of its active ingredient, chicoric acid (Jeong *et al.*, 2024). Additionally, clinical trials have demonstrated its immune-enhancing effects in adults, supporting its use as a dietary supplement (Lee *et al.*, 2024). Its adaptability to various soil types and climates further underscores its agricultural value (Mistriková and Vavrková, 2007).

A phenolic chemical, salicylic acid (SA), is an essential plant hormone and transmitter agent that controls plant development & stress tolerance (Peng *et al.*, 2021). SA, which is found naturally in very small amounts in plant tissues, is crucial for controlling several physiological and biochemical activities, including membrane permeability, ion absorption and transport, and stomatal closure. Additionally, SA is responsible for reversing the effects of ABA on leaf abscission, inhibiting ethylene production, thermogenesis, photosynthetic rate, increasing the quantity of carotenoid and chlorophyll pigments, inducing flowers, and altering the activity of some significant enzymes (Hayat & Ahmad, 2007, and Emamveridian *et al.*, 2020). Previous research on medicinal and aromatic plants has shown that SA enhances plant growth features and boosts plant production (Es-sbihi *et al.*, 2020; Gahory *et al.*, 2022; Nasseralla and Almukhtar, 2023 and El-Naggar *et al.*, 2024). Numerous studies have generally demonstrated that the usage of SA exogenously increased the effective ingredients for different varieties of aromatic & medicinal plants under unfavorable ambient conditions (Khorasani *et al.* 2023, Danesh-Shahraki *et al.* 2023, Güneri and Dalkılıç 2023).

A type of naturally occurring polysaccharide polymer found in many marine plants, including kelp and seaweed, is alginate (AA). (Liu and Zhang 2015). Alginate (ALG) is made up of heterogeneous β -1,4-glycosidic linkages connecting varying amounts of (M-blocks) D-mannuronic acid, (G-blocks) L-glucuronic acid, or both. An abundant organic material found in nature, 30,000 tons of alginate are produced annually (Donati *et al.*, 2009; Szekalska *et al.*, 2016). ALG is widely utilized in industries of food, medical, and agricultural among other sectors, as the most plentiful and inexpensive marine biomass. However, because of its large weight of molecule and poor bioavailability, alginate's uses have been severely restricted (Xing *et al.*, 2020). The novel botanical preparation has a wide range of potential uses and is decomposable, non-polluting, and comparatively safe. (Khan *et al.*, (2009). In agricultural applications, AA is frequently employed as a bioeffectors. It has the ability to stimulate plant growth. Additionally, alginate can enhance crop quality. Additionally, alginate is crucial for boosting a plant's capability to

withstand biotic and abiotic stressors (Van Oosten *et al.*, 2017).

Chitosan (Ch) is a naturally occurring natural polymer, that boosts plant growing, yield, & immunity. (Pongprayoon *et al.*, 2013; Sultana *et al.*, 2019). Even in the absence of chemical fertilizer, the use of chitosan in agriculture can greatly boost the microbial population and so convert bio-nutrients into synthetic ones that are readily taken up by the roots of plants (Bolto *et al.*, 2004). Additionally, Ch.-treated plants may be less vulnerable to ambient stressors like temperature, salt, and drought (LizarragaPauli *et al.*, 2011; Jabeen and Ahmad 2013; Pongprayoon *et al.*, 2013). Ch. is regarded globally as an environmentally safe & compatible polymer with a variety of uses in addition to a viable and affordable supplier of adaptable and efficient crop safety materials (Geisberger *et al.*, 2013; Zhang *et al.*, 2012). Ch. Hence, has a broad range of applications in several agricultural fields, such as producing crops and preservation, storage, nutritional value and so on.

One natural substitute for traditional farming methods is humic acid (HA) (Baltazar *et al.*, 2021). HA is a commonly used crop enhancer in the humic product business because it breaks down clay, buffers pH, improves holding of water and increases accessible soil nutrients (Pukalchik *et al.*, 2019, Hu *et al.*, 2021). HA It has been demonstrated that HA improves seed germination, enhances the growth of roots, leaves, and shoots, and stimulates blooming, all of which increase plant yield (Braziene *et al.*, 2021, Najarian *et al.*, 2022). HA Increased nutrient absorption, including organic matter and certain macronutrients and micronutrients, is the main cause of HA-enhanced growth. Furthermore, enhanced basis oil concentration and quality in plants like thyme, fennel, and dragonhead demonstrate how HA could profoundly alter on secondary metabolism (Noroozisharaf and Kaviani 2018; Machiani *et al.*, 2019).

This study aimed to:

Assess the effects of salicylic acid, alginate, and chitosan, either alone or in conjunction with humic acid as soil application on flowering parameters and some yield, yield components and bioactive compounds to improve these characteristics in purple coneflower *Echinacea purpurea* plant.

Methodology :-

During the two summer seasons of 2022 and 2023, two studies were conducted in the Experimental Farm Station of the Faculty of Agriculture Moshtohor, Benha Univ., Qalubia Governorate, Egypt, to study foliar application effects by salicylic acid at 200 & 400 mgL⁻¹ alginate at 250 and 500 mgL⁻¹ in addition to chitosan at 250 & 500 mgL⁻¹ with and without

humic acid soil addition compared with control (Distilled water) at flowering parameters (i.e. Number of inflorescences/plant, inflorescences FW(g)/plant. and inflorescences DW(g)/plant, and Main inflorescence height /plant, main inflorescence diameter(cm)/plant, inflorescence head FW & DW(g)/plant.), and some bio-active compounds content (i.e. (Total derivatives of caffeic acid content (mg/ g DW)) on roots, herb and flowering heads and Total alkaloids content (mg/ g DW)) in roots and areal parts), of *Echinacea purpurea* plants.

Sigma Company provided *Echinacea purpurea* seedlings that were 8–10 cm tall and had four–five leaves. In both seasons, the seedlings were transplanted into clay loam soils (**Table 1**) in the middle of April. To give the right amount of moisture for growth, the soil was immediately watered. As advised in this area, all traditional

cultural methods for cultivating *Echinacea purpurea* plants were adhered to.

The experiment treatments were as follows:

There were fourteen treatments in this study, which were the combination between humic acid soil addition 3 kg/fadden and six treatments of salicylic acid, alginate acid and chitosan as follows:

Control (Distilled water), SA at 200 mg L⁻¹, SA at 400 mg L⁻¹, AA at 250 mg L⁻¹, AA at 500 mg L⁻¹, Chit. at 250 mg L⁻¹, Chit. at 500 mg L⁻¹, Humic acid as soil application 3 kg/fadden, SA at 200 mg L⁻¹ + HA as soil application., SA at 400 mg L⁻¹ + HA as soil application., AA at 250 mg L⁻¹ + HA as soil application., AA at 500 mg L⁻¹ + HA as soil application., Chit. at 250 mg L⁻¹ + HA as soil application., Chit. at 500 mg L⁻¹ + HA as soil application.

Table 1. Physical and chemical features of the research soil throughout the 2022 as well as 2023 seasons at the Faculty of Agri., Moshtohor, Benha Univ., Qalubia Governorate.

Soil examination	Seasons	
	2022	2023
	Prior to planting	Prior to planting
Soluble cations plus anions (mmolec L⁻¹)		
Ca ⁺⁺	23.6	25.0
Mg ⁺⁺	15.0	18.0
K ⁺	3.4	3.9
Na ⁺	11.8	14.6
Cl ⁻	21.5	23.0
CO ₃ ²⁻	-	-
HCO ₃ ⁻	5.0	6.0
SO ₄ ²⁻	22.6	27.4
Chemical evaluation		
E.C (dSm ⁻¹)	6.0	5.6
PH	7.3	7.2
Soluble salts Conc. (PPM)	4800	4480
O.M (gKg ⁻¹)	1.55	1.69
CaCO ₃ (gKg ⁻¹)	1.42	1.55
Accessible N (mg/kg ⁻¹)	22.9	25
P Accessible (mg/kg ⁻¹)	9.52	11
Accessible K (mg/kg ⁻¹)	126	124
Particle size variations (Physical study)		
Course sand %	5.95	3.12
Find Sand %	23.55	22.85
Silt %	14.10	24.97
Clay %	51.25	45.10
Soil texture	Heavy Clay	Heavy Clay

Three replicates and a split-plot design were used for this investigation. Humic acid was added to the soil in the main plot. On the other hand, the subplot was used for treatments sprays i.e. control (Distilled water) salicylic acid at 200 and 400 mg L⁻¹, alginate acid at 250 and 500 mg L⁻¹. and chitosan at 250 and 500 mg L⁻¹ Through plant maturation, plants were splashed three times with various allocated solutions. The first treatment was performed 60 days following transplantation, and it continued every fifteen days.

The volume of the spraying solution was administered till plants ran off the spray solution.

Every cultural norms were carried out according advised.

Sampling and collecting data:

1. Flowering characteristics:

Three plants of echinacea from each replicates were randomly taken at 150 days after transplanting to measure plant flowering features (inflorescences number/ plant, fresh & dry weight of inflorescences,

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main inflorescence height, main inflorescence diameter & DW of main inflorescence/plant). The plant dry weight was determined after samples of the vegetative portions above ground sun-dried for 72 hours to a consistent weight.

2. Determination of some bio-active compounds content:

A- The total amount of caffeic acid derivatives in varied parts of echinacea (roots, herb and inflorescences) were measured by utilizing a spectrophotometer in accordance with **A.O.A.C (1990)**.

B- Utilizing (HPLC) High Performance Liquid Chromatography in compliance with **Bauer and Remiger (1989)**, The aerial portions of *Echinacea purpurea* (inflorescences, leaves, and stems) were used to quantify total alkaloids during both seasons (2022-2023).

Statistical analysis:

The procedures were used for statistically evaluating the study's data, outlined by **Snedecor and Cochran (1989)**. Using the statistical software program MSTAT-C (**Michigan state university, 1983**).

3. Results and discussion

1. Flowering parameters of echinacea plants:

Tables 2 and 3's data revealed that substantial differences existed between humic acid (soil addition) and without HA (soil addition) in all measurements at total flowering stage including (No. of inflorescences / plant, FW of inflorescences (g)/ plant, DW of flowering heads (g)/ plant, main inflorescence height, main inflorescence diameter, and fresh and DW of main flowering head (g)/plant) of echinacea plants across the two experimental growing seasons. In this context, applying humic acid to the soil demonstrated a notably enhanced improvement in all the traits mentioned. Conversely, soil lacking the addition of humic acid exhibited reduced values in all flowering traits across both seasons. The variations observed in the treatments analyzed could stem from the vital roles and essential functions of humic acid

These findings align with the conclusions from **Memon et al. (2014)** regarding zinnia (*Zinnia elegans*), which showed that humic acid, either alone or combined with iron sulfate, significantly enhanced the flowers number per plant, flower size (cm), Mean flower lifespan (days), and overall blooming duration (days). At the same time, the days until the first flower appeared were fewer in contrast to the control. Also, **Ahmad et al., (2019)** demonstrated that the quantity, length, and width of blooms on a plant of calendula plants increased significantly with various doses of humic acid when compared to the control.

According to Table (2), all treatments applied to the foliage (namely, salicylic acid at 200 and 400

mgL⁻¹, alginate acid at 250 and 500 mgL⁻¹, and chitosan at 250 and 500 mg L⁻¹) caused in improving all measured parameters (number of flowering heads per plant, fresh weight of flowering heads (g) per plant) to considerably rise in all measured traits (No. of inflorescences/plant, FW of inflorescences (g)/plant, DW of inflorescences (g)/plant, height of main flowering head, diameter of main flowering head, and main flowering head both fresh & dry weight(g)/plant) in contrast to the control plant. The most efficient treatment in this regard was salicylic acid at 200 mg L⁻¹, followed by chitosan acid at 500 mg L⁻¹, alginate acid at 500 mg L⁻¹, and chitosan at 250 mgL⁻¹, observed 150 days next transplanting within 2022 & 2023 seasons contrasted to untreated controls. In relation to the quantity of flowering heads per plant, there was a significant increase with most treatments applied across the two growing seasons. The only exception was a negligible increase observed with each SA @ 200 mg L⁻¹ and chitosan @ 500 mgL⁻¹ when compared to the control at 150 days post-transplanting during the 2022 and 2023 seasons, respectively. Each plant's fresh weight of inflorescences as well as its dry weight per plant displayed the highest values with chitosan at 500 mgL⁻¹, which showed the maximum values, followed by salicylic acid at 200 mgL⁻¹. In comparison with other treatments and control that yielded the minimum value within the both seasons.

Contents in Table (3) investigated that the main flowering head height/ plant and main inflorescence diameter, main inflorescence fresh and dry weight(g)/plant were recorded a significant increase with all different foliar applied treatments. Therefore, the greatest values were observed with salicylic acid & 200 mg L⁻¹ succeeded by chitosan at 500 mg L⁻¹, regarding the main inflorescence height in comparison to control during the 2022 and 2023 seasons, respectively. Conversely, the main inflorescence diameter recorded the highest values with chitosan at 500 mg L⁻¹ inflorescences salicylic acid at 200 mg L⁻¹. In contrast, the control treatment gave the lowest value during the 2022 and 2023 seasons, respectively. So, FW and DW(g) of inflorescence/ plant gave the greatest value with chitosan at 500 mg L⁻¹ as well as salicylic acid at 200 mg L⁻¹. contrasted to control through the two seasons, respectively.

Generally, it could be concluded that enhancements in inflorescences number/ plant, inflorescences weight as fresh/ plant and dry weight of inflorescences/ plant, main inflorescence height per plant, main inflorescence diameter and both the FW and DW of the main inflorescence achieved with salicylic acid at 200 mg L⁻¹. and chitosan at 500 mg/L treatments. could result from an increase in photosynthesis, leading to higher total carbohydrates and proteins in the leaves (source), as

well as an increase in endogenous auxins and other photosynthetic products, particularly during the full blooming and setting phases **Khandaker et al.; (2011) Basit et al., (2018).**

In this regard, the acquired enhancement in various growth characteristics may be ascribed to the positive impact of (Salicylic acid at 200 & 400 mgL⁻¹, Alginic acid at 250 & 500 mgL⁻¹, and chitosan at 250 & 500 mgL⁻¹) on the absorption of various minerals and additional growth substances in various plant tissues. Moreover, bio-stimulant applications lead to a rise in the number of flowers and the rate of assimilation, as well as playing crucial roles in the control of diverse vital plant processes and enhance the compounds that are involved in free radical scavenging, replacing these radicals with advantageous substances to safeguard the membranes and all organelles within the plant cell. The previously noted rise in flower quantity and the traits of flowering heads is also consistent with **Abdul Kareem and Saeed (2020) and Abdelsadek et al. (2021)**. Instead of aiding in the development of flowers, salicylic acid is involved in their commencement. SA controls floral changes during both stressful and normal circumstances by utilizing autonomous pathways and photoperiod (**Martinez et al., 2004**). offered further molecular proof of the efficacy of chitosan, which has numerous beneficial impacts on plant performance. The beneficial effects noted during the reproductive phase, like increased flowering, can also be linked to advantageous shifts in hormonal levels, improved nutrient mobilization, enhanced photosynthetic efficiency, and postponed senescence, all of which are believed to be triggered by chitosan application (**El-Sayed et al., 2020 and Abdul-Hafeez and Ibrahim 2021**).

Regarding the impact of the combination between of humic acid soil application & foliar sprays with various bio-stimulating substances, the findings submitted in the same (Tables 2 & 3) proved that the combination between HA (as drench) with SA at 200 mg L⁻¹, and HA (as drench) with Ch.at 500 mg L⁻¹ resulted in the greatest readings of each flowering traits of echinacea plants. This was succeeded by the combination treatment involving humic acid (as drench) and foliar spray with alginic acid at 500 mg L⁻¹, chitosan at 250 mg L⁻¹, salicylic acid at 400 mg L⁻¹, and alginic acid at 250 mg L⁻¹ when compared to other treatments lacking humic acid soil addition and the control in both seasons. For humic acid **Ahmed et al., (2022)** cleared that humic acid effected positively on inflorescences number/ plant, inflorescence diameter and weight as fresh and dry of inflorescences of german chamomile. As salicylic acid **Abdelsadek et al., (2021)** discovered that chrysanthemum plants sprayed with SA @ a 300

ppm resulted in early blooming (118.83 and 119.17 days post-planting). Simultaneously, applying 300 ppm SA resulted in the highest count of flower heads /plant in addition to, the inflorescence longest stalk plus thickness offlower head in chrysanthemum plant contrasted to the other applied plus control. As for chitosan **Abdul-Hafeez and Ibrahim (2021)** established that 40 ppm of chitosan yeilded the best results for various characteristics, including the flowers number for each plant FW and DW of flower heads and productivitt of dry flowers per fed compared to the untreated plants in the two seasons of the German chamomile plant. Moreover, **Moghith (2024)** reveal that chitosan foliar spray at (3 cm/L) influenced the flowering heads no. per plant, along with the FW and DW of inflorescences per plant of *Echinacea purpurea* L. in comparison to control plants.

2. Some bio-active compounds in echinacea plants:

Data presented in Tables 4 and 5 indicated that the addition of HA to soil resulted in significant differences in all chemical plant quality represented as organic components (i.e., caffeic acid derivatives and total alkamides) across both seasons. In this context, the applied of HA to the soil showed the greatest levels in roots, herbs, as well as inflorescences contents of caffeic acid derivatives, as well as total alkamides in roots and aerial parts during both seasons, in comparison to untreated soil (lacking humic acid). The variations in the examined treatments of humic acid influenced genetic factors that impacted plant quality characteristics.

In this sense, **Khorasaninejad et al. (2018)** established that the amount of derivatives of caffeic acid in the root (Cichoric acid, Chlorogenic acid, as wll as caffeic acid) significantly rose with HA soil drench compared to control on purple coneflower.

Tables 4 and 5's data revealed that chemical properties of the echinacea plant (specifically, caffeic acid derivatives and total alkamides) were significantly elevated with the applied safe agents (like SA at 200 & 400 mg L⁻¹, alginic acid at 250 & 500 mg L⁻¹, and chitosan at 250 & 500 mg L⁻¹) throughout both seasons. Echinacea plants were sprayed using SA @ 200 mg L⁻¹. revealed the largest concentration of caffeic acid compounds in the herb, roots, and inflorescences, followed by chitosan at 500 mg L⁻¹. in roots and herb. The highest value was in the inflorescences with chitosan at 250 mg L⁻¹. in comparison to control, that gave the lowest readings in caffeic acid derivatives content in root, herb and flowering heads at 150 days after transplanting during 2022 and 2023 seasons respectively as shown in **Table (4)**.

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As for alkalamides content in Table (5) echinacea plants sprayed with SA at 200 mg L⁻¹ exhibited the maximizing concentration in roots and aerial parts, while the control group showed the lowest levels for the 2022 and 2023 seasons, respectively.

In this aspect, **Mohebbi et al., (2021)** discovered that SA enhanced the levels of caffeic acid derivatives in (*Echinacea purpurea* L. Moench) when compared to the control. As for chitosan **Strzemiński et al., (2025)** found that using chitosan influences the content of secondary metabolites on *Carlina* leaves, specifically phenolic acids (chlorogenic acids), in comparison to the control. Also, **Moghith (2024)** on *Echinacea purpurea* L.

As for the interaction between humic acid (drench) and spraying with some bio-stimulants materials on chemical echinacea plant quality expressed as organic constituents (i.e., derivatives of caffeic acid, as well as total alkalamides). Findings in Tables (4 and 5) cleared that the greatest concentration in caffeic acid derivatives in root, herb and flowering heads. Total alkalamides in root and aerial parts gave maximum content as owing to

the combination of humic acid (drench) and SA at 200 mg L⁻¹ foliar spraying. during the two seasons of growth succeeded by humic acid (drench) with chitosan at 500 mg L⁻¹ as spray. These true findings throughout two experimental seasons.

These findings are in agreement with **Khorasaninejad et al., (2018)** disclosed that humic acid addition to the soil substantially raised total phenolic level in the shoots and the levels of derivatives of caffeic acid in the roots compared to a control of purple coneflower. As for salicylic acid **Mehrpooya et al., (2021)** indicated that the influence of varying salicylic acid concentrations and exposure durations, along with the 1% probability level, their interaction had an extremely substantial impact on the generation of caffeic acid derivatives in coneflowers. For chitosan **Moghith (2024)** studied that applied chitosan at (3 cm/L) had positive enhanced on Caffeic acid derivatives (8.13 and 8.28 mg/g dry weight) of *Echinacea purpurea* L. compared with control plants which gave (7.84 and 8.02 mg/g dry weight) during both seasons in succession and total alkalamides in dry aerial parts (mg/g D.W) compared with control plants.

Table 2. Impact of various applied treatments on specific floral traits (Number of inflorescences/plant, inflorescences FW(g)/plant. and inflorescences DW(g)/plant) of the echinacea plant at 150days after transplanting during the 2022 and 2023 seasons.

features		Number of inflorescences/plant.		Inflorescences fresh weight(g)/plant.		Inflorescences dry weight(g)/plant.	
		2022	2023	2022	2023	2022	2023
Treatments		150Days	150Days	150Days	150Days	150Days	150Days
A-soil application							
Without Humic		41.714 ^A	35.095 ^A	382.239 ^A	332.053 ^A	87.975 ^B	75.864 ^B
Humic acid		45.476 ^A	40.952 ^A	430.568 ^A	386.420 ^A	101.131 ^A	87.059 ^A
B – foliar spraying							
Control		21.333 ^D	17.833 ^E	155.908 ^D	135.332 ^D	27.673 ^D	24.447 ^D
Salicylic acid @ 200 mg L ⁻¹		53.000 ^A	48.167 ^A	505.845 ^{AB}	457.618 ^{AB}	120.607 ^A	99.590 ^A
Salicylic acid @ 400 mg L ⁻¹		43.333 ^{BC}	38.167 ^{CD}	412.515 ^{BC}	359.457 ^C	93.618 ^C	84.415 ^B
Alginate acid @ 250 mg L ⁻¹		40.000 ^C	35.333 ^D	364.323 ^C	322.392 ^C	88.198 ^C	75.795 ^C
Alginate acid @ 500 mg L ⁻¹		48.667 ^{AB}	42.667 ^{ABC}	456.143 ^{ABC}	405.493 ^{ABC}	106.378 ^B	98.120 ^A
Chitosan @ 250 mg L ⁻¹		46.833 ^{ABC}	39.667 ^{BCD}	433.718 ^{ABC}	373.515 ^{BC}	97.663 ^{BC}	86.650 ^B
Chitosan @ 500 mg L ⁻¹		52.000 ^A	44.333 ^{AB}	516.370 ^A	460.850 ^A	127.735 ^A	101.215 ^A
C-Interaction (A X B) (Soil application X Foliar spraying)							
Without Humic	control	17.333 ^D	12.667 ^G	122.380 ^D	109.303 ^E	17.050 ^H	14.917 ^H
	Salicylic acid @ 200 mg L ⁻¹	52.667 ^A	46.000 ^{ABC}	497.590 ^A	425.243 ^{AB}	117.200 ^{ABC}	97.010 ^{ABC}
	Salicylic acid @ 400 mg L ⁻¹	41.333 ^{BC}	35.667 ^{DE}	393.040 ^{AB}	338.650 ^{BC}	86.517 ^{EF}	77.067 ^{EF}
	Alginate acid @ 250 mg L ⁻¹	36.333 ^C	31.667 ^E	313.093 ^{BC}	279.920 ^{CD}	77.623 ^F	65.443 ^F
	Alginate acid @ 500 mg L ⁻¹	47.667 ^{AB}	40.000 ^{BCDE}	434.643 ^{AB}	383.663 ^{ABC}	102.360 ^{CDE}	95.703 ^{ABC}
	Chitosan @ 250 mg L ⁻¹	45.667 ^{ABC}	38.333 ^{CDE}	410.647 ^{AB}	357.527 ^{BC}	91.267 ^{EF}	82.087 ^{DE}
	Chitosan @ 500 mg L ⁻¹	51.000 ^{AB}	41.333 ^{BCD}	504.280 ^A	430.067 ^{AB}	123.810 ^{AB}	98.823 ^{AB}
Humic acid	control	25.333 ^D	23.000 ^F	189.437 ^{CD}	161.360 ^{DE}	38.297 ^G	33.977 ^G
	Salicylic acid @ 200 mg L ⁻¹	53.333 ^A	50.333 ^A	514.100 ^A	489.993 ^A	124.013 ^{AB}	102.170 ^{AB}
	Salicylic acid @ 400 mg L ⁻¹	45.333 ^{ABC}	40.667 ^{BCD}	431.990 ^{AB}	380.263 ^{ABC}	100.720 ^{CDE}	91.763 ^{ABCD}
	Alginate acid @ 250 mg L ⁻¹	43.667 ^{ABC}	39.000 ^{BCDE}	415.553 ^{AB}	364.863 ^{BC}	98.773 ^{DE}	86.147 ^{CDE}
	Alginate acid @ 500 mg L ⁻¹	49.667 ^{AB}	45.333 ^{ABC}	477.643 ^A	427.323 ^{AB}	110.397 ^{BCD}	100.537 ^{AB}
	Chitosan @ 250 mg L ⁻¹	48.000 ^{AB}	41.000 ^{BCD}	456.790 ^A	389.503 ^{ABC}	104.060 ^{CDE}	91.213 ^{BCD}
	Chitosan @ 500 mg L ⁻¹	53.000 ^A	47.333 ^{AB}	528.460 ^A	491.633 ^A	131.660 ^A	103.607 ^A

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Table 3. Impact of various applied treatments on specific traits flowering characteristics (Main inflorescence height /plant, main inflorescence diameter(cm)/plant, inflorescence head FW & DW(g)/plant.) of echinacea plant at 150 days after transplanting during 2022 & 2023 seasons.

Features Treatments		Main Flowering head/ plant							
		Height(cm)/plant.		Diameter(cm)/plant.		Fresh weight(g)/plant.		Dry weight(g)/plant.	
		2022	2023	2022	2023	2022	2023	2022	2023
		150Days	150Days	150Days	150Days	150Days	150Days	150Days	150Days
A-soil application									
Without Humic		62.476 ^A	56.952 ^A	4.543 ^B	4.424 ^B	20.626 ^B	19.552 ^B	4.707 ^B	4.308 ^B
Humic acid		69.167 ^A	64.762 ^A	4.895 ^A	4.743 ^A	22.326 ^A	20.930 ^A	5.264 ^A	5.022 ^A
B – foliar spraying									
Control		51.083 ^C	49.333 ^C	3.617 ^C	3.267 ^D	14.965 ^D	14.342 ^D	2.888 ^C	2.712 ^C
Salicylic acid @ 200 mg L ⁻¹		73.750 ^A	68.167 ^A	4.983 ^A	4.900 ^{AB}	24.035 ^A	22.075 ^{AB}	5.762 ^A	5.533 ^A
Salicylic acid @ 400 mg L ⁻¹		66.333 ^{AB}	62.250 ^{AB}	4.900 ^{AB}	4.750 ^{BC}	22.467 ^B	20.108 ^C	5.102 ^B	4.798 ^B
Alginate acid @ 250 mg L ⁻¹		60.250 ^B	58.667 ^B	4.683 ^B	4.667 ^C	20.632 ^C	20.080 ^C	5.033 ^B	4.565 ^B
Alginate acid @ 500 mg L ⁻¹		68.500 ^A	61.750 ^{AB}	4.900 ^{AB}	4.750 ^{BC}	22.115 ^B	21.610 ^{ABC}	5.098 ^B	4.792 ^B
Chitosan @ 250 mg L ⁻¹		69.500 ^A	60.167 ^{AB}	4.883 ^{AB}	4.800 ^{ABC}	21.673 ^{BC}	20.572 ^{BC}	5.087 ^B	4.675 ^B
Chitosan @ 500 mg L ⁻¹		71.333 ^A	65.667 ^{AB}	5.067 ^A	4.950 ^A	24.445 ^A	22.900 ^A	5.927 ^A	5.580 ^A
C-Interaction (A X B) (Soil application X Foliar spraying)									
Without Humic acid	Control	45.500 ^G	42.667 ^D	2.933 ^E	2.667 ^E	13.357 ^G	12.683 ^E	2.203 ^F	2.017 ^F
	Salicylic acid @ 200 mg L ⁻¹	70.000 ^{ABCD}	63.167 ^{ABC}	4.900 ^{ABC}	4.833 ^{ABC}	23.307 ^{ABC}	21.940 ^{AB}	5.340 ^{BC}	5.093 ^{BC}
	Salicylic acid @ 400 mg L ⁻¹	63.667 ^{CDEF}	59.500 ^{BC}	4.800 ^{ABC}	4.700 ^{BCD}	21.767 ^{CDE}	19.267 ^C	5.000 ^{CD}	4.590 ^{BCD}
	Alginate acid @ 250 mg L ⁻¹	58.500 ^{EF}	56.167 ^C	4.567 ^{CD}	4.500 ^D	19.937 ^E	19.060 ^C	4.993 ^{CD}	4.477 ^{CD}
	Alginate acid @ 500 mg L ⁻¹	66.667 ^{ABCE}	59.500 ^{BC}	4.867 ^{ABC}	4.733 ^{BCD}	22.020 ^{BCD}	21.220 ^{ABC}	5.047 ^{BCD}	4.617 ^{BCD}
	Chitosan @ 250 mg L ⁻¹	65.833 ^{BCDE}	54.833 ^C	4.767 ^{BC}	4.667 ^{CD}	20.060 ^{DE}	20.000 ^{BC}	4.890 ^D	4.207 ^D
	Chitosan @ 500 mg L ⁻¹	67.167 ^{ABCE}	62.833 ^{ABC}	4.967 ^{AB}	4.867 ^{ABC}	23.937 ^{AB}	22.697 ^A	5.473 ^B	5.157 ^B
With Humic acid	Control	56.667 ^F	56.000 ^C	4.300 ^D	3.867 ^E	16.573 ^F	16.000 ^D	3.573 ^E	3.407 ^E
	Salicylic acid @ 200 mg L ⁻¹	77.500 ^A	73.167 ^A	5.067 ^{AB}	4.967 ^{AB}	24.763 ^A	22.210 ^{AB}	6.183 ^A	5.973 ^A
	Salicylic acid @ 400 mg L ⁻¹	69.000 ^{ABCDE}	65.000 ^{ABC}	5.000 ^{AB}	4.800 ^{ABC}	23.167 ^{ABC}	20.950 ^{ABC}	5.203 ^{BCD}	5.007 ^{BC}
	Alginate acid @ 250 mg L ⁻¹	62.000 ^{DEF}	61.167 ^{ABC}	4.800 ^{ABC}	4.833 ^{ABC}	21.327 ^{CDE}	21.100 ^{ABC}	5.073 ^{BCD}	4.653 ^{BCD}
	Alginate acid @ 500 mg L ⁻¹	70.333 ^{ABCD}	64.000 ^{ABC}	4.933 ^{ABC}	4.767 ^{ABCD}	22.210 ^{BC}	22.000 ^{AB}	5.150 ^{BCD}	4.967 ^{BC}
	Chitosan @ 250 mg L ⁻¹	73.167 ^{ABC}	65.500 ^{ABC}	5.000 ^{AB}	4.933 ^{ABC}	23.287 ^{ABC}	21.143 ^{ABC}	5.283 ^{BCD}	5.143 ^B
	Chitosan @ 500 mg L ⁻¹	75.500 ^{AB}	68.500 ^{AB}	5.167 ^A	5.033 ^A	24.953 ^A	23.103 ^A	6.380 ^A	6.003 ^A

Table 4. Impact of various applied treatments on certain bio active component content (Total derivatives of caffeic acid content (mg/ g DW)) on roots, herb and flowering heads of echinacea (*Echinacea purpurea*) at 150 days after transplanting during 2022 and 2023 seasons.

Treatments	Characteristics	Total derivatives of caffeic acid content (mg/ g DW)					
		In Root		In Herb		In Flowering heads	
		2022	2023	2022	2023	2022	2023
Without humic acid	Control	5.133	5.300	7.700	6.800	9.767	8.133
	Salicylic acid @ 200 mg L⁻¹.	12.00	11.967	14.800	13.900	20.700	15.733
	Salicylic acid @ 400 mg L⁻¹.	9.400	9.267	13.800	8.610	18.500	14.233
	Alginic acid @ 250 mg L⁻¹.	7.467	7.533	13.700	9.500	13.033	12.900
	Alginic acid @ 500 mg L⁻¹.	8.633	8.400	14.200	10.533	18.200	13.033
	Chitosan @ 250 mg L⁻¹.	9.767	9.500	13.700	13.100	20.033	14.800
	Chitosan @ 500 mg L⁻¹.	10.633	11.033	14.500	13.100	18.00	14.100
With humic acid	Control	7.200	6.433	9.733	7.200	12.000	9.833
	Salicylic acid @ 200 mg L⁻¹.	14.000	12.900	17.900	17.600	21.267	17.200
	Salicylic acid @ 400 mg L⁻¹.	11.033	12.467	14.833	13.300	18.933	16.800
	Alginic acid @ 250 mg L⁻¹.	8.333	9.000	15.200	13.200	14.400	13.033
	Alginic acid @ 500 mg L⁻¹.	8.800	8.733	16.233	13.000	20.333	15.300
	Chitosan @ 250 mg L⁻¹.	11.033	10.167	15.800	13.300	21.100	16.867
	Chitosan @ 500 mg L⁻¹.	12.033	12.900	16.433	14.200	20.500	16.800

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Table 5. Impact of various applied treatments on certain bio active component content (Total alkaloids content (mg/ g DW)) in roots and areal parts of echinacea (*Echinacea purpurea*) at 150 days after transplanting during 2022 and 2023 seasons.

Treatments	Characteristics	Total alkaloids content (mg/ g dry weight)			
		In Root		In Areal parts	
		2022	2023	2022	2023
Without humic acid	Control	0.602	0.564	0.212	0.201
	Salicylic acid @ 200 mg L⁻¹.	0.814	0.785	0.41	0.385
	Salicylic acid @ 400 mg L⁻¹.	0.744	0.712	0.297	0.311
	Alginate acid @ 250 mg L⁻¹.	0.69	0.701	0.315	0.288
	Alginate acid @500 mg L⁻¹.	0.792	0.677	0.285	0.263
	Chitosan @ 250 mg L⁻¹.	0.66	0.591	0.333	0.28
	Chitosan @ 500 mg L⁻¹.	0.802	0.773	0.391	0.323
With humic acid	Control	0.698	0.69	0.23	0.219
	Salicylic acid @ 200 mg L⁻¹.	0.963	0.904	0.494	0.425
	Salicylic acid @ 400 mg L⁻¹.	0.804	0.823	0.371	0.312
	Alginate acid @ 250 mg L⁻¹.	0.797	0.800	0.398	0.35
	Alginate acid @500 mg L⁻¹.	0.911	0.872	0.401	0.344
	Chitosan @ 250 mg L⁻¹.	0.886	0.845	0.47	0.351
	Chitosan @ 500 mg L⁻¹.	0.919	0.897	0.475	0.398

References

- [1] **A.O.A.C. (1990):** Official Method of Analysis, 15th Ed., Association of official Analytical Chemists Washington DC, USA.
- [2] **Abdelsadek, O. A.; Elbohy, Naglaa, F. S. I. and Diab, Randa, I. (2021).** Impact of potassium silicate and salicylic acid on growth, flowering and quality characters of *dendranthema grandiflorum* plants under greenhouse conditions. *Scientific J. Flowers & Ornamental Plants*, 8(4):483-496.
- [3] **Abdul Kareem, A. J. and Saeed, M. (2020).** Effect of ascorbic and salicylic acids on growth and flowering of *Gazania* cv. 'Frosty Kiss Mixed'. *Ornamental Horticulture*, 26(4):537-544.
- [4] **Abdul-Hafeez, E. Y. and Ibrahim, O. H. M. (2021).** Effects of chitosan and BABA foliar application on flowering and chemical characteristics of German chamomile 'Bode-gold'. *South African J. Bot.* 139, 241-245.
- [5] **Ahmad S., Khan J., and Jamal A. (2019).** Response of pot marigold to different applied levels of humic acid. *J. Hortic. Plant Res.*, 5, 57–60.
- [6] **Ahmed, H., El-Sallami, I.; Abdul-Hafeez, E. and Ibrahim, O. (2022).** Response of growth, flowering, nutrient uptake and essential oil of German chamomile to organic nutrition. *Assiut J. Agric. Scie.*, 53. 78-92.
- [7] **Baltazar, M.; Correia, S.; Guinan, K.J.; Sujeeth, N.; Bragança, R. and Gonçalves, B. (2021).** Recent Advances in the Molecular Effects of Biostimulants in Plants: An Overview *Biomolecules*, 11, 1096.
- [8] **Basit, A.; Shah, K.; Rahman, M.; Libo X.; Zuo, X.; Han, M.; Alam, N.; Khan, F.; Ahmed, I. and Khalid, M. A. (2018).** Salicylic acid an emerging growth and flower inducing hormone in marigold (*Tagetes* sp. L.). *Pure Appl. Biol.*, 7(4): 1301-1308.
- [9] **Bauer R, and Remiger P. (1989).** TLC and HPLC Analysis of Alkamides in *Echinacea* Drugs1,2. *Planta Med.* 1989 Aug;55(4):367-71. doi: 10.1055/s-2006-962030. PMID: 17262436.
- [10] **Bolto, B.; Dixon, D. and Eldridge, R. (2004).** Ion exchange for the removal of natural organic matter. *Reactive and Functional Polymers*, 60: 171-182.
- [11] **Braziene, Z.; Paltanavicius, V. and Avizienyte, D. (2021).** The influence of fulvic acid on spring cereals and sugar beets seed germination and plant productivity *Environ. Res.*, 195, Article 110824.
- [12] **Burlou-Nagy, C.; Bănică, F.; Jurca, T.; Gratiela Vicas, L.; Marian, E.; Muresa, ME.; et al. A Pallag4-Barnes (2022).** *Echinacea purpurea* (L.) Moench: Biological and Pharmacological Properties. A Review. *Plants*. 2022; 11: 1244. <https://doi.org/10.3390/plants11091244>
- [13] **Coelho, J.; Barros, L.; Dias, M.I.; Finimundy, T.C.; Amaral, J.S.; Alves, M.J.; Calhelha, R.C.; Santos, P.F. and Ferreira, I.C.F.R. (2020).** *Echinacea purpurea* (L.) Moench: Chemical Characterization and Bioactivity of Its Extracts and Fractions. *Pharmaceuticals*, 13, 125.
- [14] **Danesh-Shahraki, H.; Ghasemi Pirbalouti, A.; Rajabzadeh, F. and Kachouei, M. A. (2023).** Foliar application of salicylic acid and proline to mitigate water deficit impact on purple coneflower (*Echinacea purpurea* (L.) Moench.). *Acta Sci. Pol. Hortorum Cultus*, 22(5), 89–97.
- [15] **Donati, I. and Paoletti, S. (2009).** Material Properties of Alginates. *Microbiology Monographs*, 1–53. doi:10.1007/978-3-540-92679-5_1
- [16] **El-Naggar, A. H. M.; Badawy, D. M.; Hassan, M. R. and Shaban, E. H. (2024).** Effect of water stress and salicylic acid on the growth and oil yield of *Ocimum basilicum* L. plant in newly reclaimed land. *Alex. Sci. Exch. J.*, 45 (1): 57-69.
- [17] **El-Sayed, Iman, M.; Salim, Rasha, G.; El-Haggar, Eman, F.; El-Ziat, Rasha, A. and Soliman, Dina, M. (2020).** Molecular characterization and positive impact of brassinosteroids and chitosan on *Solidago canadensis* cv. Tara characteristics. *Horticultrae* 6, 100.
- [18] **Emamverdian A.; Ding Y. and Mokhberdoran, F. (2020).** The role of salicylic acid and gibberellin signaling in plant responses to abiotic stress with an emphasis on heavy metals. *Plant Signaling and Behavior*, 15 (7): e1777372.
- [19] **Es-sbihi, F. Z.; Hazzoumi, Z. and Amrani, J. K. (2020).** Effect of salicylic acid foliar application on growth, glandular hairs and essential oil yield in *Salvia officinalis* L. grown under zinc stress. *Chemical and Biological Technologies in Agriculture*, 7: 1-11.

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- [20] Gahory, A. M. O.; Ayyat, A. M. and Soliman, T. M. A. (2022). Growth, yield and its component of coriander (*Coriandrum sativum* L.) in response to the addition of compost, ascorbic acid and salicylic acid under Aswan Governorate conditions, Egypt. *Journal of Plant Production*, Mansoura University, 13 (12): 899-905.
- [21] Geisberger, G.; Gyenge, E.B.; Hinger, D.; Kach, A.; Maake, C. and Patzke, G. R. (2013). Chitosan-thioglycolic acid as a versatile antimicrobial agent. *Biomacromolecules*, 14(4): 1010-1017.
- [22] Güneri, M. and Dalkılıç, Z. (2023). Effects of salicylic acid application on germination, growth and development of rough lemon (*Citrus jambhiri* Lush.) under salt stress. *Acta Sci. Pol. Hortorum Cultus*, 22, 13–26.
- [23] Gupta, M.; Sharma, D.; Sharma, A.; Kumari V. and Goshain, O.P. (2012). A review on purple coneflower (*Echinacea purpurea* L. Moench). *Journal of Pharmacy Research*, 5 (8): 4076-4081.
- [24] Hayat, S.; Ali, B. and Ahmad, A. (2007). Salicylic acid: biosynthesis, metabolism and physiological role in plant, salicylic acid. *A Plant Hormone*. Springer, Dordrecht, The Netherlands. 1-14.
- [25] Hu, Y.; Li, Q. Song, C. and Jin, X. (2021). Effect of humic acid combined with fertilizer on the improvement of saline alkali land and cotton growth. *Appl. Ecol. Environ. Res.*, 19 (2): 1279-1294
- [26] Jabeen, N. and Ahmad, R. (2013). The activity of antioxidant enzymes in response to salt stress in safflower (*Carthamus tinctorius* L.) and sunflower (*Helianthus annuus* L.) seedlings raised from seed treated with chitosan. *Journal of the Science of Food and Agriculture*, 93(7): 1699-1705.
- [27] Jeong, J.S.; Kim, J.W.; Kim, J. H.; Chung, E. H. and Lee, D.R. (2024). Oral toxicity and genotoxicity assessment of standardized *Echinacea purpurea* (L.) extract and the pharmacokinetic profile of its active ingredient chicoric acid. *Toxicological Research*, 40: 457-472.
- [28] Khan, W.; Rayirath, U. P.; Subramanian, S.; Jithesh, M. N.; Rayorath, P.; Hodges, D. M.; Critchley, A. T.; Craigie, J.S.; Norrie, J. and Prithiviraj, B. (2009). *J. Plant. Growth. Regul.* 28 386-99
- [29] Khandaker, L.; Masum Akond, A. S. M. G. and Shinya, O. B. A. (2011). Foliar application of salicylic acid improved the growth, yield and leaf's bioactive compounds in red amaranth (*Amaranthus tricolor* L.) *VEGETABLE CROPS RESEARCH BULLETIN* 74: 77-86.
- [30] Khorasani, H.; Rajabzadeh, F.; Mozafari, H. and Ghasemi Pirbalouti, A. (2023). Water deficit stress impairment of morphophysiological and phytochemical traits of *Stevia rebaudiana* Bertoni buffered by humic acid application. *S. Afr. J. Bot.*, 154, 365–371.
- [31] Khorasaninejad, S.; Ahmadabadi, A. A. and Hemmati, K. (2018). The effect of humic acid on leaf morphophysiological and phytochemical properties of *Echinacea purpurea* L. under water deficit stress. *Scientia Horticulturae* 239, 314–323
- [32] Khorasaninejad, S.; Ahmadabadi, A. A. and Hemmati, K. (2018). The effect of humic acid on leaf morphophysiological and phytochemical properties of *Echinacea purpurea* L. under water deficit stress. *Scientia Horticulturae* 239, 314–323
- [33] Lee, S.K.; Lee, D.R.; Kim, H.L.; Choi, B.K. and Kwon, K. (2024). A randomized, double-blind, placebo-controlled study on immune improvement effects of ethanolic extract of *Echinacea purpurea* (L.) Moench in Korean adults, *Phytotherapy Research*, 38 (7): 3645-3659.
- [34] Liu J F and Zhang J 2015 *Zhong Guo Guo Cai* 35(8) 20-3
- [35] Lizarraga-Pauli, E. G.; Torres-Pacheco, I.; Moreno Martinez, E. and Miranda-Castro, S. P. (2011). Chitosan application in maize (*Zea mays*) to counteract the effects of abiotic stress at seedling level. *African Journal of Biotechnology*, 10 (34): 6439-6446.
- [36] Machiani, M.A.; Rezaei Chiyaneh, E.; Javanmard, A.; Maggi, F. and Morshedloo M.R. (2019). Evaluation of common bean (*Phaseolus vulgaris* L.) seed yield and qualitative production of the essential oils from fennel (*Foeniculum vulgare* Mill.) and dragonhead (*Dracocephalum moldavica* L.) in intercropping system under humic acid application. *J. Clean.Prod.*, 235, 112-122.
- [37] Martínez, C.; Pons, E.; Prats, G. León, J. (2004). Salicylic acid regulates flowering time and links defence responses and reproductive development. *Plant J.* 37, 209-217.

- [38] Mehrpooya, Zh.; Abdoli, M. and Talebian, M. R. (2021). Effect of salicylic acid and yeast extract on caffeic acid derivatives production in *Echinacea purpurea* L. *J. Med. Plants* 20(78): 36-47.
- [39] Memon, S.A.; Bangulzai, F.M.; Keerio, M.I.; Baloch, M.A.; and Buriro, M. (2014). Effect of humic acid and iron sulphate on growth and yield of zinnia (*Zinnia elegans*). *J. Agric. Technol.*, 10(6), 1517–1529.
- [40] Michigan State university (1983): MSTAT-C: Micro-computer Statistical Program, Version 2.0. Michigan State Univ. East Lansing.
- [41] Mistríková, I. and Š. Vaverková (2007). Morphology and anatomy of *Echinacea purpurea*, *E. angustifolia*, *E. pallida* and *Parthenium integrifolium*. *Biologia*, Bratislava, 62 (1): 2-5.
- [42] Moghith, W. M. A. (2024). Influence of different irrigation intervals and anti-transpiration compounds on productivity and chemical constituents of *Echinacea purpurea* L. plant. *Egyptian J. Desert Res.*, 74, No. 2, 277-291.
- [43] Mohebbi, M.; Mortazavi, S.N.; Kheiry, A. and Saba, J. (2021). Assessment of physiological and phytochemical traits of purple coneflower (*Echinacea purpurea* L. Moench) in response to salicylic acid and methyl jasmonate application, *Medicinal Plant Research*, 11(1): 1-13.
- [44] Najarian, A.; Sour, M.K. and Nabigol, A. (2022). Influence of humic substance on vegetative growth, flowering and leaf mineral elements of *Pelargonium x hortorum*. *J. Plant Nutr.*, 45 (1): 107-112.
- [45] Nasseralla, A. A. and Almukhtar, S. A. (2023). Effect of salicylic acid and ultraviolet rays on vegetative growth characteristics of *Rosmarinus officinalis* L. in vitro. *Proceedings of the 5th International Conference for Agricultural and Environment Sciences*, 1158 (4): 8.
- [46] Noroozisharaf, A. Kaviani, M. (2018). Effect of soil application of humic acid on nutrients uptake, essential oil and chemical compositions of garden thyme (*Thymus vulgaris* L.) under greenhouse conditions. *Physiol. Mol. Biol. Pla.*, 24 (3): 423-431.
- [47] Peng, Y.; Yang, J.; Li, X. and Zhang, Y. (2021). Salicylic acid: biosynthesis and signaling *Annu. rev. plant biol.*, 72, 761-791
- [48] Pongprayoon, W.; Roytrakul, S.; Pichayangkura, R. and Chadchawan, S. (2013). The role of hydrogen peroxide in chitosan-induced resistance to osmotic stress in rice (*Oryza sativa* L.). *Plant Growth Regulation*, 70(2): 159-173.
- [49] Pukalchik, M.; Kydralieva, K.; Yakimenko, O.; Fedoseeva, E. and Terekhova, V. (2019). Outlining the potential role of humic products in modifying biological properties of the soil—A review *Front. Environ. Sci.*, 7, 80.
- [50] Salgado-Salazar, C.; Romberg, M.K. and Hudelson, B. (2023). Plasmopara echinaceae, a new species of downy mildew affecting coneflowers (*Echinacea purpurea*) in the United States. *Fungal Systematics and Evolution*, 12: 203-217.
- [51] Snedecor, G. W. and Cochran, W. G. (1989): *Statistical Method* 8th Ed., Iowa State Univ. Press. Ames, Iowa, USA.
- [52] Strzemski, M.; Dresler, S.; Hawrylak-Nowak, B.; Tkaczyk, P.; Kulinowska, M.; Feldo, M.; Maggi, F. and Hanaka, A. (2025). Chitosan elicitation enhances biomass and secondary metabolite production in *Carlina acaulis* L. *Sci Rep.* 2;15(1):23411.
- [53] Sultana, N.; Zakir, H. M.; Parvin, M. A.; Sharmin, S. and Seal, H. P. (2019). Physiological responses and nutritional qualities of tomato fruits to chitosan coating during postharvest storage. *Asian Journal of Advances in Agricultural Research*, 10(2): 1-11.
- [54] Szekalska, M.; PuciBowska, A.; Szymańska, E.; Ciosek, P. and Winnicka, K. (2016). Alginate: Current use and future perspectives in pharmaceutical and biomedical applications. *Int.J.Polym.Sci.*, 2016, 17.
- [55] Van Oosten, M.J.; Pepe, O.; De Pascale, S.; Silletti, S. and Maggio, A. (2017). The role of biostimulants and bioeffectors as alleviators of abiotic stress in crop plants. *Chem. Biol. Technol. Agric.* 4, 5. <https://doi.org/10.1186/s40538-017-0089-5>
- [56] Xing, M.; Cao, Q.; Wang, Y.; Xiao, H.; Zhao, J.; Zhang, Q.; Ji A. and Song, S. (2020). Advances in research on the bioactivity of alginate oligosaccharides. *Mar. Drugs.*, 18,144. doi:10.3390/md18030144.
- [57] Zhang, X.; Geng, X. D.; Jiang, H. J.; Li, J. R. and Huang, J. Y. (2012). Synthesis and characteristics of chitin and chitosan with the [(2-Hydroxy-3-trimethylammonium) propyl] functionality, and evaluation of their antioxidant activity in vitro. *Carbohydrate Polymers*, 89(2): 486-491.

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التأثير التكاملي بين حمض الساليسيليك، حمض الالغينيك، الشيتوزان و حمض الهيوميك لتحسين المحصول و المواد الفعالة لنبات الاخناسيا.

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أجريت تجربتان في موسمي (٢٠٢٢ و ٢٠٢٣) علي نبات الإخناسيا (*Echinacea purpurea* L.) لدراسة تأثير الرش الورقي ببعض مستحضات النمو الطبيعيه التاليه:- حمض الساليسليك بتركيز (٢٠٠ ، ٢٥٠ ملجم/لتر) وحمض الالغينيك بتركيز (٢٥٠ ، ٥٠٠ ملجم/لتر) و الشيتوزان بتركيز (٢٥٠ ، ٥٠٠ ملجم/لتر) منفردا او مع الإضافة الأرضية لحمض الهيوميك مقارنة بالكنترول رشا بالماء المقطر فقط .. علي الصفات التزهير (بداية التزهير، عدد النورات، الوزن الجاف للنورات، طول النورة الرئيسية، قطر النورة الرئيسية، الوزن الجاف للنورة الرئيسية) ومحتوي بعض المواد الفعالة للنبات (الألكاميدات و مشتقات حمض الكافيك).

ومن النتائج المتحصل عليها امكن التوصل الي :-

١- ان الرش الورقي بجميع المعاملات والتركيزات ادي الي زيادة معنوية في كلاً من صفات التزهير (بداية التزهير، عدد النورات، الوزن الجاف للنورات، طول النورة الرئيسية، قطر النورة الرئيسية، الوزن الجاف للنورة الرئيسية) وأيضاً محتوى بعض المواد الفعالة (الألكاميدات و مشتقات حمض الكافيك) في الموسمين لمقارنه بالنباتات الغير معاملة (الكنترول).

٢- اظهر الرش الورقي بكل من حمض الساليسليك بتركيز (٢٠٠ ملجم/لتر) والشيتوزان بتركيز (٥٠٠ ملجم/لتر) منفردا أو مع الاضافة الارضية لحمض الهيوميك زياده معنويه في صفات التزهير ومحتوي بعض المواد الفعالة للنبات في الموسمين متفوقا علي التركيزات الاخرى.

عموماً اكدت النتائج أن الرش الورقي بالتركيز المنخفض من حمض الساليسليك (٢٠٠ ملجم/لتر) والشيتوزان بتركيز (٥٠٠ ملجم/لتر) منفردا أو مع الاضافة الارضية لحمض الهيوميك كان أفضل المعاملات في تحسين صفات التزهير ومحتوي بعض المواد الفعالة لنبات الاخناسيا تحت ظروف الحقل وهذا ما توصي به دراسته تحت نفس الظروف.